doi: 10.25750/1995-4301-2022-1-034-043

Bioassay as a method of integral assessment for remediation of oil-contaminated ecosystems

© 2022. E. V. Morachevskaya¹ _{ORCID: 0000-0002-7176-4767}, L. P. Voronina^{1,2} _{ORCID: 0000-0003-1917-7490}, ¹Lomonosov Moscow State University, 1–12, Leninskie Gory, Moscow, Russia, 119991, ²Centre for Strategic Planning of FMBA of Russia, 10–1, Pogodinskaya St., Moscow, Russia, 119121, e-mail: Luydmila.voronina@gmail.com

The development of new technologies and improvement of existing ones for the restoration of oil-contaminated lands, the neutralization and disposal of oil drilling waste are important measures to solve environmental problems. Bioassay is successfully used to determine the danger of pollution of environmental objects by oil and oil industry wastes. With the help of biotests, it is possible to assess the state of contaminated objects, which is not always possible to do by chemicalanalytical methods, considering the complex chemical composition of petroleum hydrocarbons. An analytical review confirms the need to develop a biotesting system to assess the state of ecosystem components in the event of oil pollution and to determine the effectiveness of measures to restore them. Taking into account the nature of the pollution, presented in the article by a number of provisions on the chemical characteristics of petroleum hydrocarbons and the duration of the recovery processes, bioassay should be carried out at each stage of the reclamation process, in dynamics. Determination of the degree of neutralization of contaminated objects consists in a step-by-step transfer of the hazard level from a higher class to a lower one. Bioassay, in this case, remains a mandatory method for determining the total toxicity. In the course of sample preparation for biotesting, it is advisable to consider the possibility of increasing the bioavailability of hydrocarbon components. A prerequisite for the use of biotesting is the use of eluate and contact approaches. The main methods that can be included in the abbreviated scheme for determining the efficiency of remediation of oil-contaminated objects can be considered the method of biotesting using hydrobionts in the eluate (water extract) and phytotesting performed using the contact and eluate approaches. The battery of biological tests included in the extended scheme should be developed taking into account the specific case, taking into account the specifics of the ecosystem components, soil and climatic conditions, the methods of reclamation used, etc. The strategy for the development of biotesting is closely related to the solution of issues on the assessment of the real danger of oil pollution and the neutralization of oil drilling waste, which are among the priority ones.

Keywords: bioassay, phytoassay, drilling slurries, reclamation, hazard class.

УДК 351.777.61;632.95.024.4

Биотестирование как способ интегральной оценки приёмов рекультивации загрязнённых нефтью экосистем

© 2022. Е. В. Морачевская¹, к. б. н., в. н. с., Л. П. Воронина^{1,2}, д. б. н., в. н. с., ¹Московский государственный университет им. М. В. Ломоносова, 119991, Россия, г. Москва, Воробьёвы горы, д. 1, стр. 12, ²Центр стратегического планирования и управления медико-биологическими рисками здоровью ФМБА, 119121, Россия, г. Москва, ул. Погодинская, д. 1, стр. 1., e-mail: Luydmila.voronina@gmail.com

34

Разработка новых и совершенствование существующих технологий по восстановлению нефтезагрязнённых земель, обезвреживание и утилизация нефтебуровых отходов – важные меры по решению экологических проблем. Биотестирование успешно применяется для определения опасности загрязнения объектов окружающей среды нефтью и отходами нефтяной промышленности. С помощью биотестов можно оценить состояние загрязнённых объектов, что не всегда возможно сделать химико-аналитическими методами, учитывая сложный химический состав

нефтяных углеводородов. Аналитический обзор подтверждает необходимость разработки системы биотестирования для оценки состояния компонентов экосистемы в случае нефтезагрязнения и определения эффективности мер по их восстановлению. Принимая во внимание характер загрязнения, представленный в статье рядом положений по химической характеристике нефтяных углеводородов и длительность процессов восстановления, биотестирование следует проводить на каждом этапе процесса рекультивации, в динамике. Определение степени обезвреживания загрязнённых объектов заключается в поэтапном переводе уровня опасности из более высокого класса в более низкий. Биотестирование, в данном случае, остаётся обязательным методом определения суммарной токсичности. В ходе пробоподготовки образца для биотестирования целесообразно рассмотреть возможность увеличения биодоступности углеводородных компонентов. Обязательным условием применения биотестирования является использование элюатного и контактного подходов. Основными методами, которые могут входить в сокращённую схему определения эффективности рекультивации нефтезагрязнённых объектов, можно считать метод биотестирования с использованием гидробионтов в элюате (водная вытяжка) и фитотестирование, выполненное с применением контактного и элюатного подходов. Батарея биологических тестов, входящих в расширенную схему, должна быть разработана с учётом конкретного случая, учитывая специфику компонентов экосистемы, почвенно-климатических условий, используемых способов рекультивации и др. Стратегия развития биотестирования теснейшим образом связана с решением вопросов по оценке реальной опасности нефтяных загрязнений и обезвреживанию нефтебуровых отходов, относящихся к числу приоритетных.

Ключевые слова: биотестирование, фитотестирование, отходы нефтедобывающей промышленности, рекультивация, класс опасности, экотоксикология.

According to the official statistics of the Ministry of Energy of Russia, the level of oil production in 2020 amounted to 512.8 million tons [1]. The process of self-cleaning of natural objects takes quite a long time, especially in extreme climatic conditions [2]. In this case, a special protective function is assigned to soils, in connection with which the use of biotesting methods for assessing the integral toxicity of soil samples is an urgent issue.

Monitoring and control of the remediation process is important at all stages, and assessment of the state of oil-contaminated soils is possible using biological methods. When carrying out biotesting, it is necessary to take into account specific methodological nuances, which are not always clearly spelled out in the methodological recommendations. For example, the death of *Daphnia* in a water extract from the waste, established within 48 hours, may not exceed 10%, but the morphological state of the test object indicates a significant negative impact: individuals of *Daphnia* are smaller than in the control test, their trophic activity is minimal, and only with a longer exposure (96 h) leads to the death of the test organism [3].

The system of biotesting methods for assessing the effectiveness of methods for reclamation of soil contaminated with oil and waste from the oil industry, first of all, includes a set of biotests recommended by the regulatory document for determining the hazard class of waste (SanPiN 2.1.7.1386-03), but even for them there are nuances that associated with the specificity of the pollutant, which should be paid attention to.

The toxicity of soils, determined by the phytotesting method, directly in the soil is higher than the toxicity of aqueous extracts from the same soils [4]. The purpose of the review is to summarize the available experimental data on the effectiveness of using biotesting methods to establish toxicity in environmental objects during the remediation of oil-contaminated ecosystem components.

Objects and research methods

This review uses publications of both domestic and foreign authors (57 sources), covering the time period from 2000 to 2021. Relevance in information search was obtained by accessing the following scientific search engines: Russian scientific electronic library eLIBRARY.RU, Google scholar, Scopus/ScienceDirect. Foreign scientific electronic resources: Elsevier, Springer, ResearchGate, PubMed, Mendeley were searched by keywords and phrases: "bioassay", "drilling slurries", "reclamation", "pollution of oil products", "bioassay of oil-contaminated soils".

Chemical components of oil affecting the toxicity of oil-contaminated samples

Petroleum hydrocarbons, represented by persistent organic pollutants (POPs) and polycyclic aromatic hydrocarbons (PAHs), have a serious impact on the environment, polluting soil and water, and pose a threat to both humans and ecosystem components as a whole [5]. Of the entire list of PAHs, as a rule, only benzo[a] pyrene is standardized, which is a carcinogen and belongs to the 1st hazard class. However, this PAH has poor solubility, and chemically more active and readily soluble PAHs are not taken into account in Russian regulatory documents

[6]. Thus, the toxicity of petroleum products is determined by the hydrocarbon composition, and the toxicity of a mixture of hydrocarbons is higher than that of its individual components [7]. On the one hand, these provisions force the development of biotesting methods that allow determining possible toxic effects [8] and, on the other hand, resort to the use of sample preparation techniques for biotesting with possible preliminary dissolution of the organic compounds present, thereby increasing the toxicity index of the sample [9].

Alkanes, naphthenes and aromatic components can be processed by microorganisms, and the lighter fractions of these compounds can be completely degraded. Cyclic hydrocarbons are heavier fractions that are resistant to bacterial attack [10]. In connection with these provisions on the characterization of hydrocarbon pollution, the battery of biotests is expanding, bacterial test systems are widely used [11]. In addition to hydrocarbon pollution, during accidental oil spills, readily soluble mineral salts are often supplied to the soil, which are present in crude oil in the composition of formation waters and can also have a negative impact on biogeocenoses [12, 13].

The study of the composition of organic compounds contained in oil sludge, which is necessary to determine the toxicity of these samples [13], can be performed at the final stage, based on the results of biotesting, which will significantly reduce the time and financial costs for the chemical analysis of hydrocarbons.

Bioassay as a stage of control of efficiency of recultivation of oil-contaminated areas

In soil remediation, the following methods are used: mechanical, physicochemical, agrotechnical, microbiological, phytomeliorative [14]. The goal of reclamation is to reduce the oil content in soil and water to safe concentrations. Land reclamation should be carried out in accordance with Russian State Standard GOST R 57447-2017 in two main stages: technical and biological [15]. This indicator acts as the main and fundamental criterion after reclamation and other restoration work [16]. Determination of the mass fraction of petroleum products in soils according to the regulatory document PND F 16.1:2.2.22-98 is relevant, but the expediency of referring to this indicator is justified provided that the main volumes of the liquid fraction of petroleum products are removed, in cases of monitoring the effectiveness of reclamation, determining further stages of disposal of contaminated samples or the way of operation contaminated object. When quantitatively assessing the level of oil pollution, the fluorimetric method is widely used using the Fluorat-02 liquid analyzer (PND F 16.1: 2.21-98), as well as the methods of gas and gas-liquid chromatography [17].

Another widespread and promising reclamation technology is bioremediation. It is a process in which bacteria, fungi and plants decompose, transform and help remove pollutants while maintaining the integrity of the ecosystem. An increase in microbial density through the introduction of organic substances (compost, sewage sludge, etc.), promotes the acceleration of the decomposition of pollutants. Bioremediation can be carried out using biostimulation of aboriginal microflora or by introducing specialized microbial preparations designed to cleanse polluted ecosystems [18]. Preparations, ameliorants and materials used in this technology are preliminarily evaluated in laboratory conditions using biotest cultures [19].

Not all chemically hazardous compounds can be known, and their metabolites can be formed during biogeochemical processes [20]. At present, biotesting is widely used to control natural environments when justifying the permissible exposure to pollutants [21]. Determination of the degree of neutralization of contaminated objects consists in a step-by-step transfer of the hazard level from a higher class to a lower one [22]. Obviously, to determine the dynamics of the hazard level of oil pollution of ecosystem components, one can use biological tests included in the scheme for the experimental determination of the hazard class of waste (methods of biotesting on hydrobionts and in phytotest mentioned in the regulatory document SP 2.1.7.1386-03). According [23], most of the names of drill cuttings generated during oil production belong to the fourth class of hazard to the environment, and waste from the production of petroleum products belongs to the third class. This paradoxical information on individual components is in no way consistent with serious environmental disturbances, which once again emphasizes the relevance of choosing additional biotesting methods and revising the criteria for an adequate assessment of the state of the polluted environment (water, soil) with their help. It seems possible to use a reduced and extended biotesting scheme for an integrated assessment of the remediation of ecosystem components. The abbreviated scheme has a strictly limited number of tests (2-3 biotests). Taking into account the specifics of oil pollution, it is important that this limited set of tests be performed taking into account the necessary sample preparation and/or methodological approach: in an aqueous medium using a water extract – eluate approach; in a solid sample (for example, oil-contaminated soil) – the contact approach.

The expediency of a combination of eluate and contact biotesting

It is important to take into account that biotesting in an aqueous extract can be accompanied by significantly underestimated results in determining the degree of toxicity, while substrate biotesting allows a more complete assessment of the level of contamination. The main advantage of direct contact is that there is an interaction between the soil and the experimental test organisms, which increases the bioavailability of the contaminant. It is the use of the contact approach in biotesting increases the probability of accounting for presence of heavy metals in oil sludge increasing its toxicity. In this connection, the comparative analysis of phytotoxicity established using different approaches (eluate and applicate) differs significantly [24].

A comparative analysis of the sensitivity of two approaches (contact and eluate) in determining the toxicity of excess activated sludge of biological treatment of oily wastewater is presented [25]. Biotesting using radish (*Raphanes sativus* L.) and watercress (*Lepidium sativum* L.) seeds showed that aqueous sludge extracts at concentrations of 100, 50, and 25% were non-toxic. During testing by the contact method, directly on the sludge, a suppression of seed germination (100% phytotoxicity) was observed.

Contact tests, in contrast to eluate tests, can increase the bioavailability of pollutants and, thus, increase the toxicity index. The use of substrate bioassay, which provides direct contact of the tested organism with the test sample, allows one to establish the level of cumulative exposure from pollutants present in solid substrates [26].

The use of aquatic organisms to assess the toxicity of oil-contaminated objects

The essence of the biotesting method is to determine the effect of toxicants on specially selected test organisms with a sensitivity established under standard conditions with the registration of various behavioral, physiological, or biochemical parameters (test reactions) [27]. Although biotests fail to identify specific toxic compounds, the combination of various test systems used is indicative to determine the ecotoxicity of soils, both contaminated with oil hydrocarbons and those that have been reclaimed [9].

There is evidence that the biotesting method makes it possible to record the toxic effect of aqueous filtrates of oil-contaminated peat, even in cases where the oil content is not detected by instrumental methods (IR spectrophotometry) [28].

In cases where there is a danger of oily products entering water bodies, the use of aquatic organisms in biotesting is justified. Moreover, it should be borne in mind that along with the risk of migration of pollutants into groundwater, lateral washout into water bodies can also occur.

One of the most common test cultures used in ecotoxicological research is Daphnia magna Straus. It is a sensitive test organism that reacts sensitively to the presence of oil products of various fractions and mass concentrations in water [29]. The test response was observed in the range of concentrations of oil products from 0.012 to 200 mg/dm³ [30]. D. magna was most affected by the heaviest oil fractions of petroleum products. 100% reaction to the effects of a toxicant during the first hour from the moment of the experiment was observed at a kerosene concentration of 20 mg/dm³, oily waste -0.45 mg/dm³. Biotesting of drill cuttings and drilling mud using daphnia showed that both toxicants in the concentration range of 1.0-2.0 g/L are toxic, and only the content of 0.5 g/L in water did not have a negative effect on the vital activity of crustaceans. An increase in exposure to 25 days (chronic experiment) increased the negative effect of drill cuttings on daphnia, and at the maximum concentration for 10 days, all individuals died. By the end of the experiment, the death of daphnia at a concentration of 1.0 and 0.5 g/L was 50 and 10%, respectively [31].

The death of 50% daphnia was observed at a concentration of 0.25 mL/L of heating oil, 1.3 mL/L of diesel fuel and 5.5 mL/L of aviation gasoline. Oil was characterized by a sufficiently high toxicity for cladocerans; the death of 50% of test objects was recorded at a concentration of 0.86 mL/L. The minimum inoperative were $6.1 \cdot 10^{-3}$ mL/L of fuel oil, $3.7 \cdot 10^{-2}$ mL/L of diesel fuel, 0.53 mL/L of aviation gasoline and $1.8 \cdot 10^{-3}$ mL/L of oil [32].

In an experiment with the removal of oilcontaminated soil from lakes, an improvement in the ecological state of water bodies was observed, which affected the life of *D. magna*. After cleaning the lake, neither acute nor chronic lethal effect on *D. magna* was observed, the survival rate of crustaceans in an acute experiment (4 days) was 100%, in a chronic one (30 days) – slightly (by 10%) below the control or at the control level [33].

Based on the results of biotesting on the *D. magna* test culture, it was found that the actual level of harmless concentrations of oil products dissolved and dispersed in natural water at sampling points within the oil pollution spot on the Serebryanaya Volozhka channel (Astrakhan, Russia) is 1.4–1.5 times lower than the MPC for oil products for fishery water bodies [34].

Paramecium caudatum Ehrenberg is another crop used in the experimental determination of the hazard class of waste and for biotesting samples contaminated with oil and oil products. In a series of experiments on the impact of crude oil from the Luginetskove field, the number of freshwater ciliates of the P. caudatum species was determined in laboratory conditions for 18 days until 50% mortality was established [35]. These changes in the number of ciliates show that all the concentrations of crude oil used in the experiment (50, 100 and 200 mg/L) cause an increase in the number of paramecia on certain days of observation and reduce their number. For ciliates at a concentration of $50 \,\mathrm{mg/L}$, the mortality rate of 50% of individuals was established on 18 days of observation, at a concentration of 75 mg/L - on 12 days, with the introduction of 100 mg/L and 200 mL/L of oil on 6 and 5 days, respectively. Due to the high sensitivity, this test culture and the possibility of automatic fixation of the death of P. caudatum [36], the reproducibility of the results significantly increases, which makes this method very promising. Another indicator of toxicity when using the biotesting method with a culture of Paramecia can be the chemotaxis reaction based on the ability of ciliates to move in the direction or from the source of chemical exposure. Intact paramecium was placed in a clean drop of water connected to the second drop of a suspension of oil-contaminated soils. On day 1, the control individuals were almost evenly distributed in both drops of water; later they were concentrated in one of the drops of pure water [37].

The problem of secondary pollution often arises after the remediation of oil spill lands. The obtained results of secondary pollution showed that the adsorptive preservation of oil in the soil increases its toxicity, and this has a detrimental effect on living organisms and on the state of the soil as a whole. Changes in the survival rate of *P. caudatum* ciliates were studied at different periods of storage of samples of extracts from soils with different periods of oil pollution. It was found that with prolonged conservation of oil-contaminated soils (up to 6 months), the survival rate of ciliates decreased, and the toxicity of aqueous extracts increased [38].

It is known that one of the regularities of the toxic effect of oil pollution on protozoa is the alternation of suppression and stimulation of their biological functions. The stimulating effect of a potentially toxic substance is provided due to the mobilization of the body's reserve resources, and if the negative effect does not exceed this adaptive resource, then the stimulation can persist for a long time [39].

The test using a representative of the simplest – ciliary ciliates tetrahymen (*Tetrahymena pyriformis* Ehrenberg) is now widespread and generally accepted, convenient in cultivation and testing. Due to the fact that the ciliate is both a cell and an organism, it is possible to assess the effect of toxicants both at the cellular level and at the highest level of organization [40].

Thus, among the considered tests using aquatic organisms, especially in cases associated with oil pollution and potentially possible migration pollution of waters, cultures of *D. magna*, *P. caudatum*, *T. pyriformis* can be recommended for assessing toxicity at all stages of work on recultivation of oil-contaminated areas.

Use of higher plants for toxicity assessment of oil-contaminated facilities

One of the available methods of biotesting contaminated soils is phytotesting - diagnostics using plant organisms [41]. Phytotesting makes it possible to assess the total pollution, i. e., not only oil, but also from other by-substances that pollute the soil during oil production [42]. An express phytotest was proposed to measure the germination of seeds of white clover (Trifolium repens L.) [43]. The phytotest was developed on the example of a gray forest soil contaminated with diesel fuel or copper(II), and tested in the course of many years of experiments on the adsorption bioremediation of oil-contaminated soils. The sensitivity of the proposed method is quite high, since it can record a 10% increase in phytotoxicity of oil-contaminated soil at a concentration of the total amount of petroleum hydrocarbons of $1-5 \,\mathrm{g/kg}$, which is close to their MPC (1 g/kg) for reclaimed technogenic soils.

Considering that the germination of plant seeds in oil-contaminated soil is mainly determined by the availability of water and oxygen for

38

them, and not by the toxicity of oil [44], plants that are sensitive to this pollution are used as test cultures. For oil products and PAHs, these can be seeds of *Lactuca sativa* L., *Sorghum saccharatum* L., and *Sinapis alba* L. [27]. It was found that the rate of decrease in the content of petroleum hydrocarbons at subsequent stages of the recovery process correlates with the level of phytotoxicity, and therefore, these crops can be used to assess toxicity at all stages of bioremediation of soil contaminated with diesel fuel [21, 45].

The effect of oil on plants during soil contamination can be divided into direct toxic (stimulating) effect of hydrocarbons and other substances contained in oil, and indirect, in which changes in soil properties and transformation of the soil microbial community are possible [46]. Therefore, to assess the toxicity of samples with oil pollution, it is advisable to include in the phytotesting scheme, in addition to laboratory express methods, long-term chronic (vegetation) experiments.

In long-term experiments, when determining the degree of pollution impact on plants, a combination of factors influencing the bioavailability of pollutants is reflected. For example, as the concentration of oil increases, as well as in soil without plants, the number of saprotrophic micromycetes increases in the rhizosphere and on the surface of plant roots (rhizoplane); however, in the zone of the rhizosphere and rhizoplana this increase is more significant [47].

The discrepancies in the results of assessing the toxicity of oil by the phytotesting method can be dictated by the lack of a unified control, i. e., the changes are associated with the activity of the substrate into which this pollutant enters. Evaluation criteria should be at least two indicators: seed germination and plant growth at the juvenile stage. Germination depends on the internal energy of the plant's seed. Phytotesting methods based on the response of plants to the negative impact of pollutants are capable of providing reliable information about the quality of soils, have high sensitivity, versatility, integrity, and simplicity [41, 48].

Phytotesting carried out using the test of the culture of cress (*Lepidium sativum* L.) made it possible to establish patterns in the change in the level of pollution and the state of oil-contaminated substrate during reclamation work using sorbents [49]. Oil was added to the soil at concentrations of 1, 5, and 10% of its mass. During the entire study period, in variants with the use of sorbents, a tendency to an increase in the biometric parameters of watercress seedlings was observed. A vivid example of the effective use of phytotesting in the recovery of highly oil-contaminated soils is the results of studies of samples taken from lagoons-settling tanks using six species of higher plants: Secale cereal L., Lactuca sativa L., Zea mays L., Lepidium sativum L., Triticum vulgare L., and Brassica oleracea L. [9]. In the ecotoxicological analysis, two test parameters were investigated: seed germination and root length. The phytotesting results were compared with the results for acute toxicity according to other biotests: with luminescent bacteria Vibrio fischeri, ciliated protozoa Spirostomum ambiguum Ehrenberg, with newborn freshwater crustaceans ostracods *Heterocypris* incongruens Ramdohr. The comparability of the results, emphasize that during biogeochemical processes in the soil, more toxic metabolites or compounds can be formed.

Analysis of literature data confirms that phytotesting can be successfully used to detect oil pollution. It is important to use test cultures that are most sensitive to this type of contamination.

Use of additional analyzes to assess the toxicity of oil-contaminated objects

There is growing interest in the inclusion of several toxicity tests during remediation at the same time (with a battery of different analyzes), for a more complete ecotoxicological assessment of contaminated soils.

Such a search was carried out in an experimental work on the screening of the toxic hazard of oil-contaminated bottom sediments. in which the authors used the eluate test for *P. caudatum* in conjunction with the contact test for crustaceans Ceriodaphnia affinis Lilljeborg and Hyalella azteca Saussure [50]. A group of authors has developed a test system consisting of the microbial strain Pseudomonas putida [51]. In another work [52], it is proposed to use the gram-positive bacterium Bacillus pumilus KM-21 as a test culture to determine the toxicity of HM and the possibility of this method for assessing the toxicity of oil waste. The authors of [53] propose to use three test objects for ecotoxicological assessment: hydrobionts (D. magna), soil microorganisms (soil respiration), and higher plants (Avena sativa). In world practice, an integral estimate is widely used, calculated taking into account the results of a set of methods used the Triad approach [54]. This approach, along with biotesting, which reflects the ecotoxicological characteristics, includes methods of chemical analysis and bioindication parameters taking into account the biological activity and response of microbial communities.

The promising application of biotesting soils and water bodies contaminated with oil and oil products not only with the help of higher plants, but also with the help of a bioluminescent test for photobacteria [55]. The use of these methods, which determine the reducing activity of microorganisms in conjunction with growth tests on bacteria and microalgae, reflects the pollution of water and soil with oil products. The sensitivity of contact and eluate tests using microbial test organisms V. fischeri (bioluminescence inhibition test) and Azomonas agilis Beijerinck (dehydrogenase activity test) has also been reported [56].

The toxicity found in tests based on the use of direct contact with the sample (applicate testing methods) correlates more closely with the results of physicochemical analyzes. Currently, it is the "tests of direct contact" with the soil, for example, with the earthworms *Eisenia foetida* Michaelsen, the standard test GOST 33036-2014, in which bioindicators are in close contact with toxic substances adsorbed on soil particles, that are quite promising. Soil pollution with oil has a long-term negative effect on soil animals, causing their almost complete elimination in the obligate zone of pollution and a sharp decline in numbers even with weak pollution. The toxic effect of oil on earthworms is determined not only by the intensity of pollution, but also by the morpho-ecological characteristics of certain species and their belonging to different natural and climatic zones [57].

Bioassaying aquatic organisms and higher plants to assess oil pollution are well-studied, but not the only methods. The use of bacteria as test organisms is currently being actively studied due to the fact that these test organisms are highly sensitive to the pollutants under study and grow well in laboratory conditions. These data can be used to create mandatory methods for biotesting oil-contaminated objects and are included in the biotest system, in which the presence of representatives of all the main kingdoms of the living is important: bacteria, fungi, plants and animals.

Conclusions

Despite the fact that there is no universal test system for the determination of all existing toxicants, biotesting as a method of integral assessment is successfully used as a tool for environmental monitoring. Biotesting has a good potential for evaluating the effectiveness of technologies for cleaning and restoring oilcontaminated components of the ecosystem, which is advisable to carry out in dynamics.

Taking into account the low bioavailability of pollutants (hydrophobic organic toxicants), it is advisable to use eluate and contact biotesting approaches as complementary to each other. To improve the accuracy of biotesting results in the course of determining the toxicity of heterogeneous oily components in combination with other toxicants, the possibility of using sample preparation reagents for dissolving/leaching poorly soluble organic compounds should be considered.

The biotesting system can be represented by reduced (basic version) and extended schemes. To select the battery of biotests used in the abbreviated scheme, it is possible to propose methods using aquatic organisms and plants, which are recommended in environmental and hygienic legislation. Hydrobionts D. magna Straus or *P. caudatum* Ehrenberg are sensitive test organisms and are able to diagnose both acute and chronic toxicity of aqueous extracts of oil-contaminated objects. Higher plants L. sativa L., S. saccharatum L., A. sativa L., and S. alba L. are widely tested and are used as a test culture for phytotesting oil-contaminated soils. These biotesting methods comply with Russian state standards and methodological recommendations, have a clear execution protocol and are used to assess the effectiveness of reclamation techniques.

The extended research scheme is carried out taking into account the specifics of the contaminated objects, reclamation measures and according to the results obtained in the abbreviated scheme. The extended scheme covers a wide range of tested organisms and their reaction levels: higher plants, soil biota, protozoa, hydrobionts, microorganisms, luminescent bacterial tests, biochemical enzymatic methods, etc. Selection of a complex of sensitive and physiologically different test cultures, determination of informative test indicators, the choice of highly reproducible methods, a summary approach to effective assessment, processing of results, automation of methods are the necessary conditions for the biotesting system to assess the effectiveness of reclamation measures.

To develop a unified strategy for classifying oil pollutants and determining their real hazard, scientific institutions need to establish communication with regulatory organs that are involved in the prevention of environmental hazards.

40

Acknowledgments: The reported study was funded by RFBR, project number 20-14-50312.

References

1. Ministry of Energy of the Russian Federation [Internet resource] https://minenergo.gov.ru/node/910 (Accessed: 16.07.2021) (in Russian).

2. Pushkareva M.V., Mai I.V., Seredin V.V., Leybovich L.O., Chirkova A.A., Vekovshinina S.A. Environmental assessment of the surrounding area and population health state in the oil production areas in Perm region // Environmental protection in oil and gas complex. 2013. No. 2. P. 40–45 (in Russian).

3. Olkova A.S. Properties and problems of biotesting of aqueous media using certified methods // Voda: khimiya i ekologiya. 2014. No. 10. P. 87–94 (in Russian).

4. Bakina L.G., Bardina T.V., Mayachkina N.V. Methodic approaches of phytoassay of technogenically polluted soils // Environmental problems of the northern regions and ways to solve them: Materialy Mezhdunarodnoy konferentsii. 2004. Part. 1. P. 167–169 (in Russian).

5. Sammarco P.W., Kolian S.R., Warby R.A.F., Bouldin J.L., Subra W.A., Porter S.A. Concentrations in human blood of petroleum hydrocarbons associated with the BP/Deepwater Horizon oil spill, Gulf of Mexico // Arch. Toxicol. 2016. V. 90. P. 829–837. doi: 10.1007/s00204-015-1526-5

6. Khaustov A.P., Redina M.M. Geochemical markers based on concentration ratios of PAH in oils and oil-polluted areas // Geochemistry International. 2017. No. 1. P. 57-67 (in Russian).

7. Smolnikova V.V., Yemelyanov S.A., Dementyev M.S. Influence of oil hydrocarbons on the environment and ways of sanitation the petropolluted substrata // Izvestiya Samarskogo nauchnogo tsentra Rossiyskoy akademii. 2009. V. 11. No. 1 (6). P. 1378–1380 (in Russian).

8. Leme D.M., Marin-Morales M.A. *Allium cepa* test in environmental monitoring: a review on its application // Mutation Research. 2009. V. 682. No. 1. P. 71–81. doi: 10.1016/j.mrrev.2009.06.002

9. Płaza G., Nałecz-Jawecki G., Ulfig K., Brigmon R.L. The application of bioassays as indicators of petroleum-contaminated soil remediation // Chemosphere. 2005. V. 59. No. 2. P. 289–296. doi: 10.1016/j.chemosphere.2004.11.049

10. Nazarko M.D., Shcherbakov V.G., Aleksandrova A.V. Prospects for the use of microorganisms for biodegradation of oil-contaminated soils // Izvestiya VUZov. Pishchevaya tekhnologiya. 2004. No. 4. P. 89–91 (in Russian).

11. Juvonen R., Martikainen E., Schultz E., Joutti A., Ahtiainen J., Lehtokari M. A battery of toxicity tests as indicators of decontamination in composting oily waste // Ecotoxicology and Environmental Safety. 2000. V. 47. No. 2. P. 156–166. doi: 10.1006/eesa.2000.1943 12. Avetov N.A., Arzamazova A.V., Kinzhaev R.R., Trofimov S.Ya., Shishkonakova E.A. Methodic approaches to assessment of oil-polluted lands in middle Priobye region, West Siberia // Problemy agrohimii i ekologii. 2012. No. 3. P. 56–61 (In Russian).

13. Voronina L.P., Ponogaybo K.E. An approach to the choice of bioassay methods for soil research // Agrochemistry. 2021. No. 9. P. 77–81 (in Russian).

14. Bobrenko E., Yugov Ya. Specific aspects of oilcontaminated land reclamation at the oil and gas field // Elektronnyy nauchno-metodicheskiy zhurnal Omskogo GAU. 2018. No. 3 (14). P. 1–13. [Internet resource] http://e-journal.omgau.ru/images/issues/2018/3/00601. pdf (Accessed: 16.07.2021) (in Russian).

15. Rogova N.S. Causes of the inefficient reclamation of oil-contaminated lands in Western Siberia // Geoecology. 2020. No. 1. P. 82–85 (in Russian).

16. Chuksin I.V. Problems of recultivation of oil-polluted lands // Nauchno-prakticheskiy elektronnyy zhurnal Alley of science. 2020. No. 6 (45). P. 176–181 (in Russian).

17. Kuznetsova T.O. Methods of assessment and control of oil pollution of soils education // Science-production: Materialy Vserossiyskoy nauchno-prakticheskoy konferentsii. 2018. V. 2. P. 122–127 (in Russian).

18. Masciandaro G., Macci C., Peruzzi E., Ceccanti B., Doni S. Organic matter-microorganism-plant in soil bioremediation: a synergic approach // Rev Environ Sci Biotechnol. 2013. V. 12. P. 399–419. doi: 10.1007/s11157-013-9313-3

19. Timanov A.S., Ashikhmina T.Ya., Leshchenko A.A., Pogorelskiy I.P., Sharov S.A., Teterin B.B., Lazykin A.G., Filimonova G.V., Ezhov A.V., Permyakov R.G. A biological product with an extended spectrum of biodegradative activity for soil reclamation of the chemical weapons destruction facility "Maradykovsky" // Theoretical and Applied Ecology. 2015. No. 3. P. 61–69 (in Russian). doi: 10.25750/1995-4301-2015-3-042-061-069

20. Shen W., Zhu N., Cui J., Wang H., Dang Z., Wu P., Luo Y., Shi C. Ecotoxicity monitoring and bioindicator screening of oil-contaminated soil during bioremediation // Ecotoxicol Environ Saf. 2016. V. 124. P. 120–128. doi: 10.1016/j.ecoenv.2015.10.005

21. Arzamazova A.V., Kinzhaev R.R., Trofimov S.Ya. Application of spring wheat in phytotesting of oil-polluted soils // Problemy agrohimii i ekologii. 2016. No. 2. P. 47–51 (in Russian).

22. Pozdniakov S.A., Kesler K.E., Voronina L.P. The approach to studying and neutralization petroleum sludge as a part of utilization of sludge pits // Wastes, the reasons for their formation and prospects for use: Sbornik nauchnykh trudov po materialam Mezhdunarodnoy nauchnoy ekologicheskoy konferentsii. Krasnodar: Kubanskiy gosudarstvennyy agrarnyy universitet imeni I.T. Trubilina, 2019. P. 337–339 (in Russian).

23. Federal Classification Catalog of Waste [Internet resource] https://docs.cntd.ru/document/542600531 (Accessed: 03.05.2021) (in Russian).

24. Pukalchik M.A., Terekhova V.A., Karpukhin M.M., Vavilova V.M. Comparison of eluate and direct soil bioassay methods of soil assessment in the case of contamination with heavy metals // Eurasian Soil Science. 2019. No. 4. P. 507–514 (in Russian).

25. Nafikova L.F., Trubnikova L.I., Erokhina N.I., Bogdanova A.R. Phytotoxicity of the sludge of biological treatment of oily wastewater // Aspirant. 2017. No. 5. P. 88–92 (in Russian).

26. Boży M. Assessment of phytotoxicity of leachates from landfilled waste and dust from foundry // Ecotoxicology. 2020. V. 29. P. 429–443. doi: 10.1007/s10646-020-02197-1

27. Terekhova V.A., Voronina L.P., Kiryushina A.P., Morachevskaya E.V., Kydralieva K.A. Standard algorithm for measuring phytoeffects. Moskva: Scientific Library of the Lomonosov Moscow State University, 2021. 58 p. (In Russian).

28. Kudryavtsev A.A., Mikhailova L.V., Rybina G.E., Gordeeva F.V., Tsulaiya A.M., Znamenzhikov A.N. Study of migration and degradation of oil in peat soil of upland bogs in KHMAO under control of biotesting // Sibirskiy Ekologicheskiy Zhurnal (Siberian Journal of Ecology). 2012. No. 6. P. 761–768 (in Russian).

29. Sipulinov R.B., Karagaycheva Yu.V., Shilova N.A., Rogacheva S.M. Estimation the toxicity of oil production waste by biotesting methods // Izvestiya Samarskogo nauchnogo tsentra Rossiyskoy akademii nauk. 2015. V. 17. No. 5 (2). P. 695–699 (in Russian)

30. Fomicheva G.P., Kamakin A.M., Fyodorova I.V. Determining the degree of toxicity of natural surface water polluted with oil products by methods numerical chemical analysis and bio-testing // Vestnik AGTU. Ser.: Rybnoe khozyaystvo. 2016. No. 4. P. 42–49 (in Russian).

31. Kryuchkov V.N., Kurapov A.A. Assessment of the impact of drilling waste on hydrobionts // Vestnik AGTU. Ser.: Rybnoe khozyaystvo. 2012. No. 1. P. 60–65 (in Russian).

32. Lozovoy D.V., Dagurov A.V., Potapov D.S., Stom D.I. Estimation of toxity of mixtures of non-iongenic surfaceactive substances and oil products for *Daphnia* // Byulleten VSNTs SO RAMN. 2005. No. 6 (44). P. 146–149 (in Russian).

33. Belskaya M.A., Rybina G.E. Assessment of the toxicity of bottom soils of some lakes of the Nizhnevartovsky district using *Daphnia magna* Straus // The youth's view on solving problems of the development of the agroindustrial complex in the conditions of globalization in modern society. Sbornik materialov XLIX Mezhdunarodnoy studencheskoy nauchno-prakticheskoy konferentsii. 2015. Part I. P. 155–158 (in Russian).

34. Fomicheva G.P., Nasibulina B.M., Biryukova M.G. Crustaceans of the Volga delta in emergency conditions of oil spills // Vestnik AGTU. Ser.: Rybnoe khozyaystvo. 2020. No. 1. P. 27–34 (in Russian).

35. Izeneva S.V. Investigation of oil pollution impacts for freshwater infusions *Paramecium caudatum //*

Sovremennye issledovaniya. 2018. No. 5 (9). P. 251–253 (in Russian).

36. Mai T.L., Voronina L.P., Cheremnikh E.G. Bioassay of soils on rice fields in Vietnam // Moscow University Bulletin. Series 17. Soil science. 2014. No. 3. P. 28–35 (in Russian).

37. Gordeeva F.V., Mikhaylova L.V., Petukhova G.A. Vital parameters *Paramecium caudatum* in water extracts of oil polluted soil // Tyumen State University Herald. 2009. No. 3. P. 232–237 (in Russian).

38. Artemenko S., Vanyuhova A. Biotesting secondary petropollutid adsorbed by soil // Tyumen State University Herald. 2017. V. 3. No. 4. P. 66–73 (in Russian).

39. Gordeeva F.V. Results of biotesting of bottom deposits of some lakes in Khanty-Mansi autonomous okrug – Yugra // Vestnik rybokhozyaystvennoy nauki. 2017. V. 4. No. 4 (16). P. 87–93 (in Russian).

40. Valiullina A.M. Biotesting as a method of environmental monitoring // Nauchno-prakticheskie issledovaniya. 2020. No. 8–3 (31). P. 32–34 (in Russian).

41. Voronina L.P. Plant bioassay for environmental assessment of agrocenosis // Problemy agrokhimii i ekologii. 2013. No. 3. P. 16–21 (in Russian).

42. Klimova A.A, Azarova S.V, Yazikov E.G, Matveenko A. Scanning electron microscopy study of drilling cuttings in Tomsk oblast sites // IOP Conf. Series: Earth and Environmental Science. 2019. P. 1–7. doi: 10.1088/1755-1315/272/2/022004

43. Vasilyeva G.K., Kondrashina V.S., Strijakova E.R., Pinsky D.L. Express-phytotest for choosing conditions and following process of soil remediation // Environ Geochem Health. 2020. Sep 26. doi: 10.1007/s10653-020-00727-8

44. Nazarov A.V. Use of microbial-plant associations for bioremediation of oil contaminated soil // Izvestiya Samarskogo nauchnogo tsentra Rossiyskoy Akademii nauk. 2013. V. 15. No. 3–5. P. 1673–1675 (in Russian).

45. Ovsyannikova I.V., Shulaev N.S., Pryanichnikova V.V., Kadyrov R.R. Determination of toxicity of soil contaminated with by of petrochemical hidrocarbons // Journal of Physics Conference Series. 2021. V. 1889. No. 3. Article No. 032011. doi: 10.1088/1742-6596/1889/3/032011

46. Nazarov A.V. The effect of oil pollution of the soil on plants // Bulletin of Perm University. Biology. 2007. V. 5. No. 10. P. 134–141 (in Russian).

47. Sakaeva E.Kh., Rudakova L.V. Assessment of the biological activity of industrial soils based on drill slurries for reclamation of disturbed lands // Theoretical and Applied Ecology. 2020. No. 4. P.192–197 (in Russian). doi: 10.25750/1995-4301-2020-4-192-197

48. Voronina L.P., Terekhova V.A., Morachevskaya E.V. Phytoassay in environmental control. Moskva: Scientific Library of the Lomonosov Moscow State University, 2021. 34 p. (in Russian).

49. Fomina N.V. Phytotesting of oil polluted soilgrants after use of sorbents // Epokha nauki. 2017. No. 11. P. 154–161 (in Russian). doi: 10.1555/2409-3203-2017-0-11-154-161

42

50. Stepanova N.Yu., Akhmetshina A.D., Latypova V.Z. Test-organism sensitivity comparison in toxicological evaluation of bottom sediments polluted with oil of various origin // Povolzhskiy Journal of Ecology. 2012. No. 3. P. 319–325 (In Russian).

51. Selivanovskaya S.Yu., Latypova V.Z. Development of a test system for assessing the toxicity of multicomponent wastes stored in the natural environment // Russian Journal of Ecology. 2004. No. 1. P. 21–25 (in Russian).

52. Selivanovskaya S.Yu., Galitskaya P.Yu., Latypova V.Z., Semanov D.A. Evaluation of efficiency of contact and eluate methods of biotesting of soils // Uchenye Zapiski Kazanskogo Universiteta. Seriya Estestvennye Nauki. 2007. V. 149. No. 1. P. 113–122 (in Russian).

53. Mayachkina N.V., Chugunova M.V. Peculiarities of soil biotests to evaluate soil ecotoxicity // Vestnik of Lobachevsky University of Nizhni Novgorod. 2009. No. 1. P. 84–93 (in Russian). 54. ISO 19204:2017. Soil quality – Procedure for site-specific ecological risk assessment of soil contamination (soil quality TRIAD approach) [Internet resource] https://www.iso.org/standard/63989.html (Accessed: 02.06.2021).

55. Palamae S., Sompongchaiyakul P., Suttinun O. Effects of crude oil and aromatic compounds on growth and bioluminescence of *Vibrio campbellii* FS5 // Environ Monit Assess. 2021. V. 193. Article No. 291. doi: 10.1007/s10661-021-09081-3

56. Leitgib L., Kálmán J., Gruiz K. Comparison of bioassays by testing whole soil and their water extract from contaminated sites // Chemosphere. 2007. V. 66. No. 3. P. 428–34. doi: 10.1016/j.chemosphere.2006.06.024

57. Kibardim V.M., Artiemieva T.I., Zherebtzov A.K. Effects of oil pollution on earthworms in different natural and climatic zones // Uchenye Zapiski Kazanskogo Gosudarstvennogo Universiteta. 2008. V. 150. No. 1. P. 97–105 (in Russian).